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# *In vitro* **Antioxidant/ Radical Scavenging Activities and Hepatoprotective Roles of Ethanolic Extract of** *Cassia occidentalis* **Leaves in Sodium Arsenite-Treated Male Wistar Rats**

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# *Authors' contributions*

*This work was carried out in collaboration between the authors. Author MAG designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Author OAO contributed to the literature searches and corrected the draft manuscript. All authors read and approved the final manuscript.*

*Research Article*

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# **ABSTRACT**

**Aims**: To investigate *in vitro* antioxidant/radical scavenging activities and hepatoprotective ability of ethanolic leaf extracts of *Cassia occidentalis* (COLEX) in male Wistar rats treated with sodium arsenite (NaAsO<sub>2</sub>).

**Study Design/Methodologies**: Using four different methodologies, the anti-oxidant/free radical scavenging activities of COLEX were determined in comparison with standard antioxidants, butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT). For the hepatoprotection study, four groups of rats were used. Groups **A**: Control group given distilled water only; **B**: Given NaAsO<sub>2</sub> at 2.5 mg•kg $^{-1}$  bw/day (p.o.) for 2 weeks; **C**:  $\vert$ Administered COLEX alone at 200 mg•kg<sup>-T</sup> bw a day for 2 weeks (p.o.); **D**: Pre-treated | with COLEX for 2 weeks followed by  $N_{\text{AASO}_2}$ . The activities of the enzymes aspartate and alanine aminotransferases (AST and ALT), alkaline phosphatase (ALP) and γ glutamyl transferase (γGT) were determined in the treated and control animals as indices of hepatotoxicity.

**Place and Duration of the Study**: The animal treatment and analyses were carried out at Department of Biochemistry, University of Ibadan between February and June 2008.

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**Results**: At 25, 40 and 50  $\mu$ g•m<sup>-1</sup> concentrations of the extracts or the antioxidants, the reducing power is of the order BHA > BHT > COLEX. At 50  $\mu$ g•ml<sup>-1</sup>, the percentage inhibitions of peroxidation by COLEX, BHA and BHT were respectively 96.2%, 97.3% and 98.4% while percentage DPPH scavenging effect of COLEX, BHA and BHT were 62.5%, 67.5% and 61.3% respectively. The  $H_2O_2$  scavenging activities were respectively 53.0%, 85.3% and 97.8% for COLEX, BHA and BHT. Pre-treatment with COLEX before administration of NaAsO<sub>2</sub> led to significant ( $p < 0.05$ ) reduction in the mean liver and serum γGT, and serum ALP and AST activities when compared with group administered only NaAsO<sub>2</sub>.

**Conclusion**: COLEX exhibited hepatoprotective effects against NaAsO<sub>2</sub> toxicity in male rats.

*Keywords: Cassia occidentalis; hepatoprotection; antioxidant; sodium arsenite; carbon tetrachloride; γ-glutamyl transferase.*

# **ABBREVIATIONS**

*γGT: γ-glutamyl transferase; ALP: Alkaline phosphatase); BHA: Butylated hydroxyanisole; BHT: Butylated hydroxytoluene; COLEX: Ethanolic extracts of Cassia occidentalis leaves.*

## **1. INTRODUCTION**

Human population is exposed to a number of chemicals either accidentally, occupationally or through life style habits. Many of these foreign substances exhibit their toxic effects by generating reactive oxygen species and other free radicals which cause damages to cells and various cellular molecules. The reactive oxygen species are causative factors in the aetiologies of many diseases such as cancers and hepatopathies [1-3]. The crucial role of the liver in the metabolism of foreign chemical substances in the body makes it one of the organs evoking toxic responses and an important target for toxicological studies.

A significant environmental contaminant is sodium arsenite. It is a component of herbicides, fungicides, insecticides, algaecides and arsenical soap [4-7]. Drinking water with high inorganic arsenic concentration is the main source of arsenic exposure for the world population [8]. Many studies have proved sodium arsenite to be highly toxic [9-11]. Other studies have shown that sodium arsenite is hepatotoxic and clastogenic in experimental animals [12-14].

Dietary intervention and the use of natural products, as found in the medicinal plants, offer possible means of control of the incidence of food/contaminants related diseases [15-16]. *Cassia occidentalis* L is a plant that has been widely employed in indigenous and folk medicine in the treatment of a wide range of ailments such as antidote of poison, blood purifier, expectorant, anti-inflammatory agent and a remedy for the treatment of liver diseases [17-18]. It is used in various liver conditions such as anaemia, hepatitis and liver damage. It could possibly help prevent liver cells damage. Water-ethanolic extract of Nigerian *C. occidentalis* leaf have been shown to contain alkaloids, tannins, saponins and phlobatannins [19]. Ethnobotany, phytochemical, pharmacological and toxicological activities of *C. occidentalis* have been comprehensive reviewed by Yadav et al. [20]. In the present study, we investigated (i) *in vitro* antioxidant/ radical scavenging activities of ethanolic extracts of Cassia occidentalis leaves (COLEX) in comparison with standard antioxidants,

butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT); and (ii) its hepatoprotective ability of male Wistar rats treated with sodium arsenite.

# **2. MATERIALS AND METHODS**

# **2.1 Chemicals**

Sodium arsenite, BHA (butylated hydroxyanisole), BHT (butylated hydroxytoluene) were purchased from Sigma Chemical Co., St Louis, MO, USA. Kits for ALT, AST, ALP and γGT were purchased from Randox Laboratories Ltd (UK). All other reagents and chemicals were of analytical grade, products of Sigma Chemical Co, or BDH Chemical Ltd, Poole, England.

# **2.2** *Cassia occidentalis* **and Extraction Procedures**

Fresh *Cassia occidentalis* leaves were collected from the Botanical Garden, University of Ibadan and were authenticated at the Herbarium, Department of Botany, University of Ibadan. The leaves were dried at room temperatures on the laboratory bench surface. The dried leaves were ground into a fine powder with an electric blender and extracted with 70% ethanol solution. Extraction methods used were according to Gulcin [21]. Thereafter, the solvent were removed using rotary evaporator. The extract was placed in a plastic bottle and stored at -20ºC until used.

# **2.3 Antioxidant and Radical Scavenging Activities of** *Cassia occidentalis* **Leaves Extract**

## **2.3.1 Total reductive capacity**

The reductive capacity of COLEX was determined by the method of Oyaizu [22]. This method was described comprehensively by Gulcin et al. [21]. Briefly, to 1 ml of COLEX, BHA and BHT (each at 25, 40 and 50  $\mu$ g•ml<sup>-1</sup>), were added 2.5 ml phosphate buffer (0.2 M, pH 6.6) and 2.5 ml of 1% potassium ferricyanide,  $[K_3Fe(CN)_6]$ . The mixture was incubated at 50ºC for 20 min. 2.5 ml of 10% trichloroacetic acid were added to each of the mixture which was then centrifuged at 1000 x g. The upper layer of the mixture was pipetted out and mixed with 2.5 ml distilled water and 0.5 ml of 0.1% FeCl<sub>3</sub>. Absorbance was measured at 700 nm in a Spectrumlab 23A spectrophotometer (Techmel & Techmel Texas USA). Increased absorbance of the reaction mixture indicates increased reducing power.

## **2.3.2 Total antioxidant activity determination by ferric thiocyanate method**

The method of Mitsuda et al. [23] with the modifications described by Gulcin et al. [24] was employed for the determination of total antioxidant activity of ethanolic extract of *Cassia occidentalis* leaves (COLEX) and standards antioxidant compounds (BHA and BHT). For each, a stock solution of 1 mg•m $^{-1}$  distilled water was prepared. Then, each of the solution containing different concentrations (25, 40 and 50  $\mu$ g•ml<sup>-1</sup>) of the extract or the standards in 2.5 ml of potassium phosphate buffer (0.04 M, pH 7.0) was added to 2.5 ml of linoleic acid emulsion. The linoleic emulsion (5 ml) contained 17.5 µg Tween 20, 15.5 µl linoleic acid and 0.04 M potassium phosphate buffer. On the other hand, the control was composed of 2.5 ml potassium phosphate buffer and 2.5 ml of linoleic acid emulsion. The mixed solutions were incubated at 37ºC in a glass flask in the dark. The peroxide levels were determined by reading the absorbance at 500 nm in a Spectrumlab 23A spectrophotometer (Techmel &

Techmel Texas USA) after reaction with  $FeCl<sub>2</sub>$  and thiocyanate at intervals during incubation. These steps were repeated until the control reached its maximum absorbance value. The percentage inhibition of lipid peroxidation in linoleic acid emulsion was calculated by the following:

Inhibition of lipid peroxidation (%) = 100 -  $\left[\frac{A_{\text{Control}} - A_{\text{Sample}}}{A_{\text{Control}}}\right]$  X 100

Where  $A_{sample}$  is the absorbance in the presence of the extract COLEX or standard compound BHA or BHT and A<sub>control</sub> is the absorbance of the control reaction mix.

## **2.3.3 1,1-Diphenyl-2-picryl-hydrazyl (DPPH) free radical scavenging activity**

The scavenging capacity of free radical (DPPH) by COLEX, BHA and BHT were evaluated as described by Elmastas [25]. This involves the monitoring of the bleaching rate of the stable free radical DPPH at the characteristic wavelength (517 nm) in the presence or absence of the sample. An antioxidant would decrease DPPH absorbance [25]. A 1 ml solution (0.1 mM) of DPPH in ethanol was added to 3 ml of different concentrations (25, 40 and 50 µg/ml) of COLEX, BHA or BHT. The absorbance was read at 517 nm after 30 minutes. A lower absorbance of the reaction mixture indicates higher free radical scavenging activity. The capacity to scavenge DPPH radical was calculated as follows:

**DPPH** scavenging effect (%) = 
$$
\left(\frac{A_{\text{Control}} - A_{\text{Sample}}}{A_{\text{Control}}}\right) \times 100
$$

Where  $A_{Control}$  is the absorbance of the control reaction and  $A_{Sample}$  is the absorbance in the presence of the sample (COLEX, BHA or BHT).

#### **2.3.4 Hydrogen peroxide scavenging activity**

The hydrogen peroxide scavenging ability of COLEX was determined according to the method of Ruch et al. [26]. A 40 mM solution of  $H_2O_2$  was prepared in phosphate buffer (pH 7.4). Extracts or standard (50  $\mu$ g•ml<sup>-1</sup>) was added to 0.6 ml of the H<sub>2</sub>O<sub>2</sub>. The absorbance of the mixture was read at 230 nm. Blank solution contained the buffer without  $H_2O_2$ . The percentage  $H_2O_2$  scavenged by the COLEX, BHA and BHT was calculated as follows:

H<sub>2</sub>O<sub>2</sub> scavenged activity (%) = 
$$
\left[\begin{array}{c} A_{\text{Control}} - A_{\text{Sample}} \\ \hline \\ A_{\text{Control}} \end{array}\right] \times 100
$$

Where  $A_{\text{Control}}$  is the absorbance of the control reaction and  $A_{\text{Sample}}$  is the absorbance in the presence of the sample (COLEX, BHA or BHT).

# **2.4 Experimental Animals and Treatments**

Male Wistar rats approximately thirteen weeks old and in the weight range of (130 – 180) g, healthy and with no injury were obtained from the Animal House of the Faculty of Basic Medical Sciences. They were housed in steel metal cages in the Animal House of Department of Biochemistry. The rats were divided into four groups (A - D) of 5 animals each, based on the treatment received. *Cassia occidentalis* leaves extract was suspended in convenient volume of distilled water and administered as indicated below.

- **A** : Group of rats given distilled water only.
- **B** : Animals in this group were given sodium arsenite at 2.5 mg•kg<sup>-1</sup> body weight/day (p.o.) for 2 weeks.
- **C** : Animals in this group were given *Cassia occidentalis* leaf extract alone by oral at 200 mg•kg<sup>-1</sup> body weight a day for 2 weeks by oral intubation.<br>**D**: This group were pre-treated with *Cassia occidentalis* leaf extract (p.o.) at 200 mg•kg
- <sup>1</sup> body weight/day for 2 weeks followed by sodium arsenite at 2.5 mg•kg<sup>-1</sup> body weight for 2 weeks.

All the rats were fed rodent pellets and given drinking water *ad libitum*. The rats were allowed to acclimatise for seven days before the commencement of the experiment and kept at 12 hours light/ dark cycle and temperature of  $29+2$ °C throughout the duration of the experiment. Following the above treatment, the animals were fasted 24 hours before they were sacrificed by cervical dislocation.

## **2.4.1 Enzyme assays**

From each of the sacrificed animals, serum was separated from blood that has been allowed to clot at room temperature (for about two hours). The clotted blood samples were centrifuged at 3,000 x g for 10 minutes; the supernatant (i.e. the serum) was separated and used immediately or stored at -20ºC until required. Livers were washed in phosphate buffered saline, blotted dry on filter paper and weighed. The liver was homogenized in 4 volume of phosphate buffered saline. The homogenate was centrifuged at 12,000 x g for 20 minutes and the supernatant stored at -20ºC.

## **2.4.2 Gamma-glutamyl transferase (γGT) activity**

γGT was assayed in the serum and liver homogenates by using the reconstituted γGT diagnostic reagent following the method of Szasz [27].

#### **2.4.3 Alanine aminotransferase (ALT) and aspartate aminotransferase(AST) activities**

Serum ALT and AST were assayed according to Reitman and Frankel [28]. This method involves the reaction of pyruvate, the product of transamination reaction catalysed by ALT or AST, with 2, 4-dinitrophenyl hydrazine to produce intensely coloured hydrazone read at 546 nm using spectronic-20 spectrophotometer.

## **2.4.4 Alkaline phosphatase (ALP) activity**

Serum ALP assay was based on the method of Williamson [29]. This involves a spectrophotometric (405 nm) determination of concentration of p-nitrophenol formed by the dephosphorylation of p-nitrophenyl phosphate (PNPP) catalysed by ALP.

## **2.4.5 Statistical analysis**

The data were analysed by One-way Analysis of Variance (ANOVA) followed by analysis of Least Significant Difference (LSD). Values were considered statistically significant when *P*=0.05. All the results are expressed as mean + standard deviation.

## **3. RESULTS**

## **3.1 Antioxidant and Radical Scavenging Activities of** *Cassia occidentalis* **Leaves Extract**

Total reducing capability, total antioxidant activity, DPPH radical and hydrogen peroxide scavenging assays were used for the evaluation of antioxidant activities of ethanolic extract of *Cassia occidentalis* leaves. It was found that the reducing ability of the COLEX and standard compounds, BHA and BHT increased with increasing concentrations. At all concentrations, the reducing power is in the order BHA > BHT > COLEX (Fig. 1). Secondly, the total antioxidant activity of COLEX, BHA and BHT were determined by ferric thiocyanate in linoleic acid emulsion method. There was increase absorbance at 500 nm with the increasing concentration of COLEX, BHA and BHT from  $25 - 50 \mu g \cdot m l^{-1}$  as shown in Fig. 2. The corresponding percentage inhibition of peroxidation in the linoleic acid system (calculated as in material and methods section) was found to increase steadily for COLEX, BHA and BHT with increasing concentration (Fig. 3). At 50  $\mu$ g•ml<sup>-1</sup>, the percentage inhibitions of peroxidation by COLEX, BHA and BHT were respectively 96.2%, 97.3% and 98.4% (Fig. 3). Furthermore, the antioxidant activities of COLEX, BHA and BHT at different concentrations were also determined by DPPH scavenging method. There were increased DPPH scavenging abilities with increasing concentration of COLEX and the standard compounds (Fig. 4). At 50  $\mu$ g•ml<sup>-1</sup> concentration, the calculated percentage scavenging effect of the extract, COLEX (62.5%), is similar to those of BHA and BHT (67.5% and 61.3 % respectively). Likewise, the hydrogen peroxide scavenging activities of COLEX, BHA and BHT were determined at 50  $\mu$ g•ml<sup>-1</sup> concentration in each case. The H<sub>2</sub>O<sub>2</sub> scavenging activities were respectively 53.0%, 85.3% and 97.8% for COLEX, BHA and BHT (Fig. 5).

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**Fig. 1. Total reductive capability of COLEX, BHA and BHT at different concentrations using the potassium ferricyanide reduction method. COLEX is ethanolic extracts of** *Cassia occidentalis* **leaves; BHA (butylated hydroxyanisole); BHT (butylated hydroxytoluene). Indicated are the mean values for duplicate analyses**



**Fig. 2. Total antioxidant activities at different concentrations of COLEX, BHA and BHT using ferric thiocyanate in linoleic acid emulsion method. COLEX is ethanolic extracts of** *Cassia occidentalis* **leaves; BHA (Butylated hydroxyanisole); BHT (Butylated hydroxytoluene). Indicated are the mean values for duplicate experiments**

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**Fig. 3. Total antioxidant activities of COLEX, BHA and BHT at different concentrations measured as % lipid peroxidation inhibition. COLEX is ethanolic extracts of** *Cassia occidentalis* **leaves; BHA (Butylated hydroxyanisole); BHT (Butylated hydroxytoluene). Indicated are the mean values for duplicate analyses**



**Fig. 4. Free radical (DPPH) scavenging ability of COLEX, BHA and BHT at different concentrations. COLEX is ethanolic extracts of** *Cassia occidentalis* **leaves; BHA (Butylated hydroxyanisole); BHT (Butylated hydroxytoluene). Values are means for duplicate analyses**





## **3.2 Hepatotoxicity of Sodium Arsenite: Protective Effects of** *Cassia occidentalis* **Extracts**

Experimental rats administered sodium arsenite (group B) recorded mean γ-glutamyl transferase (γGT) and alkaline phosphatase (ALP) activities that were significantly (p < 0.05) greater than the observation made with the negative control (group A) given distilled water (Figs. 6 and 7). Pre-treatment of the animals with extract of *C. occidentalis* resulted in significant (p < 0.05) reduction of the effect of sodium arsenite on the γGT and ALP activities (Figs. 6 and 7). Similarly rats administered sodium arsenite alone (group B) have elevated serum alanine and aspartate aminotransferase activities that are significantly ( $p < 0.05$ ) different from the respective values obtained with the negative control group (Fig. 8). Treatments of the animals with ethanolic extract of *C. occidentalis* before the administration of sodium arsenite resulted in the attenuation of the sodium arsenite induction of these serum enzymes (group D) to levels that are significantly ( $p < 0.05$ ) lower than group given sodium arsenite alone. However, the observed enzyme activities made with animals pretreated with the *C. occidentalis* extract before the toxin are in all cases significantly (p < 0.05) higher that the negative control group (group D vs A, Figs. 6, 7 and 8).



#### **Fig. 6. Effect of ethanolic extracts of** *Cassia occidentalis* **leaves (COLEX) on sodium arsenite induction of liver and serum γ-glutamyl transferase (γGT)**

A: Control group of rats given distilled water only. B: Group of rats given sodium arsenite at 2.5 mg·kg<sup>-1</sup> body weight/day (p.o.) for 2 weeks (positive control); **C**: animals in this group were given Cassia *occidentalis leaf extract alone at 200 mg•kg-1 body weight a day for 2 weeks by oral intubation D: This group were pre-treated with Cassia occidentalis leaf extract at 200 mg•kg-1 body weight/day for 2 weeks followed by sodium arsenite at 2.5 mg•kg-1 body weight for 2 weeks. Values are Mean + SD; n=5. \*Significantly different from control group A (p < 0.05). #Significantly (p < 0.05) different from positive control group B.*



#### **Fig. 7. Effect of ethanolic extracts of** *Cassia occidentalis* **leaves (COLEX) on sodium arsenite induction of serum alkaline phosphatase**

A: Control group of rats given distilled water only. B: Group of rats given sodium arsenite at 2.5 mg·kg<sup>-1</sup> body weight/day (p.o) for 2 weeks (positive control); C: animals in this group were given Cassia *occidentalis leaf extract alone at 200 mg•kg-1 body weight a day for 2 weeks by oral intubation D: This group were pre-treated with Cassia occidentalis leaf extract at 200 mg•kg-1 body weight/day for 2 weeks followed by sodium arsenite at 2.5 mg•kg-1 body weight for 2 weeks. Values are Mean + SD; n=5. \*Significantly different from control group A (p < 0.05). #Significantly (p < 0.05) different from positive control group B.*



**Fig. 8. Effect of ethanolic extracts of** *Cassia occidentalis* **leaves (COLEX) on sodium arsenite induction of serum amino transferases (ALT and AST)**

A: Control group of rats given distilled water only. B: Group of rats given sodium arsenite at 2.5 mg·kg<sup>-1</sup> body weight/day (p.o) for 2 weeks (positive control); C: animals in this group were given Cassia *occidentalis leaf extract alone at 200 mg•kg-1 body weight a day for 2 weeks by oral intubation D: This group were pre-treated with Cassia occidentalis leaf extract at 200 mg•kg-1 body weight/day for 2 weeks followed by sodium arsenite at 2.5 mg•kg-1 body weight for 2 weeks. Values are Mean + SD; n=5. \*Significantly different from control group A (p < 0.05). #Significantly different (p < 0.05) from positive control group B.*

# **4. DISCUSSION**

Free radicals are causative factors in the aetiologies of many diseases [2-3]. They are easily generated through normal metabolic pathways in the body. Antioxidants, on the other hand are able to quench free radicals thereby preventing diseases. Natural antioxidants of plant sources have been associated with reduction of chronic diseases through their capacity to terminate free radical propagation in biological systems [30]. This capacity is widely used as a parameter to characterise nutritional and medicinal values of plants and their bioactive components.

In the present study, we have determined the antioxidant and free radical scavenging activities of ethanolic extract of *Cassia occidentalis* leaves (COLEX) in comparison with the standard antioxidants, BHA and BHT using four different methodologies including total reducing capability, total antioxidant activity, DPPH radical and hydrogen peroxide scavenging assays. In addition, we assessed the effect of COLEX on the sodium arseniteinduced hepatotoxicity in the male Wistar rats.

Potassium ferricyanide reduction method was used to determine the reducing power of COLEX, BHA and BHT. The reducing power of a compound serves as an indicator of its potential antioxidant activity. The reduction of  $Fe^{3+i}$ ferricyanide complex to the ferrous form is proportional to the power of the antioxidant substances present in the samples. The  $Fe<sup>2+</sup>$ 

formed as Perl's Prussian blue is then monitored at 700 nm [31]. The reducing ability of the samples is concentration dependent, increases with increasing concentration of the extract and the standards. At 50  $\mu$ g•ml<sup>-1</sup> concentration, the reductive power of COLEX is approximately 69% of BHT and 48% of BHA values (Fig. 1). The total antioxidant activity of *Cassia occidentalis* leaves extract, COLEX and the reference compounds, BHA and BHT (determined by percentage inhibition of lipid peroxidation) with ferric thiocyante method showed that the extract compared very well with the reference compounds (Fig. 3). The percentage inhibition calculated were of the order BHT > BHA > COLEX.

However, the percentage DPPH scavenging effect of COLEX, BHA and BHT were found to be very similar. A close look at the results of this assay indicates that COLEX was more efficient than BHT in scavenging DPPH radicals while BHA superseded COLEX as DPPH scavengers (Fig. 4). On the other hand, assay of hydrogen peroxide scavenging activities proved BHA and BHT to be more efficient than COLEX (Fig. 5). Aqueous extract of leaves of *C. occidentalis* have been shown to be most effective as free radical scanvenger in comparison with methanolic, chloroform, petroleum ether and benzene extracts [32]. The extensive antioxidant/ free radical activities of COLEX could be linked with the phytochemicals such as alkaloids, tannins, sapponins and phlobatannins identified in the water-ethanolic extract of Nigerian *C. occidentalis* leaves [19]. Moreover, recent studies have demonstrated that the phenolic compounds were responsible for the antioxidant potency of leaf and stem barks extracts of *C. occidentalis* and some medicinal plants [32- 33]. Antioxidant properties, most especially free radical scavenging abilities, of components of food sources are very important due to the deleterious role of free radicals in food and in biological systems. Excessive generation of free radical accelerates the oxidation of lipids and other macromolecules in foods thereby decreasing food quality and consumer acceptance [34-37]. In addition, free radicals have been linked with many chronic pathological conditions [38-44]. On the other hand, effective food sourced antioxidant systems are able to prevent the above deleterious processes.

Short term exposure to sodium arsenite has been shown to induce hepatotoxicity in experimental animals [12-13,45] but mechanism of its hepatotoxicity is still under investigation. In the present study, we found experimental rats treated with sodium arsenite (group B) have mean liver and serum γGT, serum ALP, AST and ALT activities that are significantly higher ( $p < 0.05$ ) than the value observed for the negative control group (group A). This is consistent with findings with other hepatotoxins [46-47]. In other words, the elevation in the level of these enzymes is an indication of liver lesion [48-50]. On the other hand, we found that the ethanolic leaf extracts of *C. occidentalis* attenuates the sodium arsenite-induced enzymes. Pre-treatment with leaf extracts of *C. occidentalis* before administration of sodium arsenite led to significant ( $p < 0.05$ ) reduction in the mean liver and serum γGT and serum ALP and AST activities when compared with groups administered only sodium arsenite, supporting the presence of hepatoprotective components in the extracts against the hepatotoxicity of sodium arsenite.

Hepatotoxicity of sodium arsenite has been linked with its ability to generate free radicals *in vivo* [14,51-52]. It is possible that COLEX is able protect against sodium arsenite induced hepatotoxicity in the experimental animals because of its antioxidant and free radical scavenging activities. The findings here give scientific support and allude to the indigenous and folk medicine use of *Cassia occidentalis* L as a remedy for the treatment of liver diseases [17-18]. These findings are also in line with reports on extracts from other plants in rendering protective effects against chemicals and toxins [53-55].

## **5. CONCLUSION**

Findings from this study revealed that ethanolic extract of *Cassia occidentalis* leaves possesses potent antioxidant and free radical scavenging activities. The extract also exhibited protective effects against sodium arsenite-induced hepatotoxicity in male rats.

# **CONSENT**

Not applicable.

# **ETHICAL APPROVAL**

All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985), as well as University of Ibadan ethics committee rules on the use of laboratory animals in research were followed.

# **COMPETING INTERESTS**

Authors have declared that there are no competing interests.

# **REFERENCES**

- 1. Malins DC, Holmes EH, Polissar NL, Gunselman SJ. The etiology of breast cancer. Characteristic alteration in hydroxyl radical-induced DNA base lesions during oncogenesis with potential for evaluating incidence risk. Cancer. 1993;71(10):3036-43.
- 2. Poli G. Liver damage due to free radicals. Br Med Bull. 1993;49(3):604-20.
- 3. Beckman KB, Ames BN. The free radical theory of aging matures. Physiol Rev. 1998;78(2):547-81.
- 4. Griffon H, Kiger J, Delabesse P. Comparative analytical study of various commercial shampoos and possible consequences of their use concerning the content of arsenic of the hair. Ann Pharm Fr. 1961;19:407-18.
- 5. Da Costa EW. Variation in the toxicity of arsenic compounds to microorganisms and the suppression of the inhibitory effects by phosphate. Appl Microbiol. 1972;23(1):46- 53.
- 6. Shariatpanahi M, Anderson AC, Abdelghani AA, Englande AJ, Hughes J, Wilkinson RF. Biotransformation of the pesticide sodium arsenate. J Environ Sci Health B. 1981;16(1):35-47.
- 7. Chen Z, Cai Y, Solo-Gabriele H, Snyder GH, Cisar JL. Interactions of arsenic and the dissolved substances derived from turf soils. Environ Sci Technol. 2006;40(15):4659- 65.
- 8. Chiou HY, Hsueh YM, Liaw KF, Horng SF, Chiang MH, Pu YS, et al. Incidence of internal cancers and ingested inorganic arsenic: a seven-year follow-up study in Taiwan. Cancer Res. 1995;55(6):1296-300.
- 9. Biswas R, Poddar S, Mukherjee A. Investigation on the genotoxic effects of long-term administration of sodium arsenite in bone marrow and testicular cells in vivo using the comet assay. J Environ Pathol Toxicol Oncol. 2007;26(1):29-37.
- 10. Yousef MI, El-Demerdash FM, Radwan FM. Sodium arsenite induced biochemical perturbations in rats: ameliorating effect of curcumin. Food Chem Toxicol. 2008;46(11):3506-11.
- 11. El-Demerdash FM, Yousef MI, Radwan FM. Ameliorating effect of curcumin on sodium arsenite-induced oxidative damage and lipid peroxidation in different rat organs. Food Chem Toxicol. 2009;47(1):249-54.
- 12. Odunola OA, Uka E, Akinwumi KA, Gbadegesin MA, Osifeso OO, Ibegbu MD. Exposure of laboratory mice to domestic cooking gas: implications for toxicity. Int J Environ Res Public Health. 2008;5(3):172-6.
- 13. Gbadegesin MA, Odunola OA, Akinwumi KA, Osifeso OO. Comparative hepatotoxicity and clastogenicity of sodium arsenite and three petroleum products in experimental Swiss Albino Mice: the modulatory effects of Aloe vera gel. Food Chem Toxicol. 2009;47(10):2454-7.
- 14. Chattopadhyay S, Deb B, Maiti S. Hepatoprotective role of vitamin B(12) and folic acid in arsenic intoxicated rats. Drug Chem Toxicol. 2012;35(1):81-8.
- 15. Wattenberg LW. Chemoprevention of cancer. Cancer Res. 1985;45(1):1-8.
- 16. Popkin BM. Understanding global nutrition dynamics as a step towards controlling cancer incidence. Nat Rev Cancer. 2007;7(1):61-7.
- 17. Nadkarni AK. Indian Materia Medica. Bombay: Popular publication; 1976. 289 pp..
- 18. Chopra RN, Nayar SL, Chopra IC. Glossary of Medicinal plants. New Delhi: CSIR; 1980. 55 pp.
- 19. Ogunkunle ATJ, Ladejobi TA. Ethnobotanical and phytochemical studies on some species of Senna in Nigeria. Afr J Biotechnol 2006;5: 2020–3.
- 20. Yadav JP, Arya V, Yadav S, Panghal M, Kumar S, Dhankhar S. *Cassia occidentalis* L.: a review on its ethnobotany, phytochemical and pharmacological profile. Fitoterapia. 2010;81(4):223-30.
- 21. Gulcin I, Elmastas M, Aboul-Enein HY. Determination of antioxidant and radical scavenging activity of Basil (*Ocimum basilicum* L. Family Lamiaceae) assayed by different methodologies. Phytother Res. 2007;21(4):354-61.
- 22. Oyaizu M. Studies on product of browning reaction prepared from glucose amine. Jpn J Nutr 1986;44:307– 315.
- 23. Mitsuda H, Yuasumoto K, Iwami K. Antioxidation action of indole compounds during the autoxidation of linoleic acid. Eiyo to Shokuryo 1996;19:210-214.
- 24. Gulcin I. The antioxidant and radical scavenging activities of black pepper (Piper nigrum) seeds. Int J Food Sci Nutr. 2005;56(7):491-9.
- 25. Elmastas M, Turkekul I, Ozturk L, Gulcin I, Isildak O, Aboul-Enein HY. Antioxidant activity of two wild edible mushrooms (Morchella vulgaris and Morchella esculanta) from North Turkey. Comb Chem High Throughput Screen. 2006l;9(6):443-8.
- 26. Ruch RJ, Cheng SJ, Klaunig JE. Prevention of cytotoxicity and inhibition of intercellular communication by antioxidant catechins isolated from Chinese green tea. Carcinogenesis. 1989;10(6):1003-8.
- 27. Szasz G. A kinetic photometric method for serum gamma-glutamyl transpeptidase. Clin Chem. 1969;15(2):124-36.
- 28. Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. Am J Clin Pathol. 1957l;28(1):56-63.
- 29. Williamson T. A comparison between the phosphastrate and phenyl phosphate methods of alkaline phosphatase assay. Med Lab Technol. 1972;29(2):182-7.
- 30. Covacci V, Torsello A, Palozza P, Sgambato A, Romano G, Boninsegna A, Cittadini A, Wolf FI. DNA oxidative damage during differentiation of HL-60 human promyelocytic leukemia cells. Chem Res Toxicol. 2001;14:1492–7
- 31. Chung YC, Chang CT, Chao WW, Lin CF, Chou ST. Antioxidative activity and safety of the 50 ethanolic extract from red bean fermented by Bacillus subtilis IMR-NK1. J Agric Food Chem. 2002;50(8):2454-8.
- 32. Arya V, Yadav S, Kumar S, Yadav JP. Antioxidant activity of organic and aqueous leaf extracts of *Cassia occidentalis* L. in relation to their phenolic content. Nat Prod Res. 2011;25(15):1473-9.
- 33. Choudhary RK, Swarnkar PL. Antioxidant activity of phenolic and flavonoid compounds in some medicinal plants of India. Nat Prod Res. 2011;25(11):1101-9.
- 34. Tsikunib AD, Iudina TV, Istomin AV, Klepikov OV. Medical and biological problems of relationship between food product quality and free radical oxidation state in humans. Vopr Pitan. 2000;69(5):28-31.
- 35. Stanley DW. Biological membrane deterioration and associated quality losses in food tissues. Crit Rev Food Sci Nutr. 1991;30(5):487-553.
- 36. Addis PB. Occurrence of lipid oxidation products in foods. Food Chem Toxicol. 1986;24(10-11):1021-30.
- 37. Wojciak KM, Dolatowski ZJ. Oxidative stability of fermented meat products. Acta Sci Pol Technol Aliment. 2012;11(2):99-109.
- 38. Rossman MJ, Groot HJ, Reese V, Zhao J, Amann M, Richardson RS. Oxidative Stress and COPD: The Impact of Oral Antioxidants on Skeletal Muscle Fatigue. Med Sci Sports Exerc; 2013 Jan 4.
- 39. Condezo-Hoyos L, Rubio M, Arribas SM, Espana-Caparros G, Rodriguez-Rodriguez P, Mujica-Pacheco E, et al. A plasma oxidative stress global index in early stages of chronic venous insufficiency. J Vasc Surg. 2013;57(1):205-13.
- 40. Ferrari CK, Torres EA. Biochemical pharmacology of functional foods and prevention of chronic diseases of aging. Biomed Pharmacother. 2003;57(5-6):251-60.
- 41. Pontiki E, Kontogiorgis C, Xu Y, Hadjipavlou-Litina D, Luo Y. New lipoxygenase inhibitors of reactive oxygen species production in cellular models of amyloid (A2) toxicities. J Alzheimers Dis. 2013;34(1):215-30.
- 42. Pollack RL, Morse DR. Free radicals and antioxidants: relation to chronic diseases and aging. Int J Psychosom. 1988;35(1-4):43-8.
- 43. Basso D, Panozzo MP, Fabris C, del Favero G, Meggiato T, Fogar P, et al. Oxygen derived free radicals in patients with chronic pancreatic and other digestive diseases. J Clin Pathol. 1990;43(5):403-5.
- 44. Rahman I, Tomasi A. 2nd International Meeting on Free Radicals in Health and Disease. The role of oxidants and antioxidants in the regulation of chronic diseases, may 8-12, 2002, Istanbul, Turkey. Free Radic Res. 2003;37(4):349-54.
- 45. Wijeweera JB, Thomas CM, Gandolfi AJ, Brendel K. Sodium arsenite and heat shock induce stress proteins in precision-cut rat liver slices. Toxicology. 1995;104(1-3):35- 45.
- 46. Hsiao G, Lin YH, Lin CH, Chou DS, Lin WC, Sheu JR. The protective effects of PMC against chronic carbon tetrachloride-induced hepatotoxicity in vivo. Biol Pharm Bull. 2001;24(11):1271-6.
- 47. Ritter C, Reinke A, Andrades M, Martins MR, Rocha J, Menna-Barreto S, et al. Protective effect of N-acetylcysteine and deferoxamine on carbon tetrachlorideinduced acute hepatic failure in rats. Crit Care Med. 2004;32(10):2079-83.
- 48. Ketterer B, Meyer DJ. Glutathione transferases: A possible role in the detoxification oand repair of DNA and lipid hydroperoxides. Mutat Res. 1989;45:1-8.
- 49. Lum G, Gambino SR. Serum gamma-glutamyl transpeptidase activity as an indicator of disease of liver, pancreas, or bone. Clin Chem. 1972;18(4):358-62.
- 50. Ideo G, Morganti A, Dioguardi N. Gamma-glutamyl transpeptidase: a clinical and experimental study. Digestion. 1972;5(6):326-36.
- 51. Chattopadhyay S, Maiti S, Maji G, Deb B, Pan B, Ghosh D. Protective role of Moringa oleifera (Sajina) seed on arsenic-induced hepatocellular degeneration in female albino rats. Biol Trace Elem Res. 2011;142(2):200-12.
- 52. Bashir S, Sharma Y, Irshad M, Nag TC, Tiwari M, Kabra M, et al. Arsenic induced apoptosis in rat liver following repeated 60 days exposure. Toxicology. 2006;217(1):63-70.
- 53. Biswas S, Talukder G, Sharma A. Protection against cytotoxic effects of arsenic by dietary supplementation with crude extract of Emblica officinalis fruit. Phytother Res. 1999;13(6):513-6.
- 54. Karthikeyan K, Ravichandran P, Govindasamy S. Chemopreventive effect of Ocimum sanctum on DMBA-induced hamster buccal pouch carcinogenesis. Oral Oncol. 1999;35(1):112-9.
- 55. Rastogi S, Shukla Y, Paul BN, Chowdhuri DK, Khanna SK, Das M. Protective effect of Ocimum sanctum on 3-methylcholanthrene, 7,12-dimethylbenz(a)anthracene and aflatoxin B1 induced skin tumorigenesis in mice. Toxicol Appl Pharmacol. 2007;224(3):228-40.

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